



# Just Natural Science<sup>™</sup> La science au naturel, simplement<sup>™</sup>

# VimForte-VM<sup>™</sup>

VimForte-VM<sup>™</sup> is a nutritional supplement of macro-minerals, micro-minerals, fat-soluble vitamins, water-soluble vitamins and antioxidants essential for diverse biochemical functions.

#### **INDICATIONS** For the treatment and/or prevention of vitamin and mineral deficiencies associated with:

• Chronic diseases • Convalescence

Malabsorption

- Infections
- Postoperative • Preoperative
- Restricted diets • Stress

### **INGREDIENTS** • Antioxidant

- **ACTIONS** Enzyme Cofactors
  - Growth
  - Energetic
  - Metabolic
  - Restorative

# **VimForte-VM<sup>™</sup> FORMULA**

1 teaspoon (5 mL) contains:

Cholecalciferol	(Vitamin D3)	200 IU	Calcium	(Citrate)	50 mg
d-alpha-Tocopherol	(Vitamin E)	50 IU	Magnesium	(Citrate)	30 mg
beta-Carotene		2500 IU	Boron	(Sodium Borate)	500 mcg
Thiamine Hydrochloride	(Vitamin B1)	2 mg	Chromium	(Polynicotinate)	10 mcg
Riboflavin	(Vitamin B2)	2 mg	Copper	(Gluconate)	0.5 mg
Niacinamide	(Vitamin B3)	15 mg	Manganese	(Sulphate)	2 mg
Calcium Pantothenate	(Vitamin B5)	5 mg	Selenium	(Sodium Selenite)	25 mcg
Pyridoxine Hydrochloride	(Vitamin B6)	1 mg	Silicon	(Sodium Metasilicate)	5 mg
Biotin	(Vitamin B7)	25 mcg	Zinc	(Citrate)	5 mg
Folic Acid	(Vitamin B9)	100 mcg	<b>Choline Bitartrate</b>		10 mg
Cyanocobalamin	(Vitamin B12)	25 mcg	Inositol		35 mg
Ascorbic Acid	(Vitamin C)	50 mg	Lecithin		25 mg

## **ADMINISTRATION** Oral

**DOSAGE** 1 - 20 lbs 1.25 ml (<sup>1</sup>/<sub>4</sub> teaspoon) daily.

AphaVet Science

VimForte-VM

500 mL

Multivitamin and Mineral Fo

- 21-50 lbs  $2.5 \text{ ml} (\frac{1}{2} \text{ teaspoon}) \text{ daily.}$
- 51-100 lbs 5 ml (1 teaspoon) daily.
- > 100 lbs 10 ml (2 teaspoons) daily.
- **STORAGE** Refrigerate after opening. Keep bottle cap tightly closed when not in use. Keep out of reach of children.
- **PACKAGING** 500 mL/bottle

#### NON-MEDICINAL INGREDIENTS

Stevia, Citric acid, Potassium sorbate, Purified water, Sodium benzoate.



VimForte-VM<sup>TM</sup> is manufactured under strict GMP standards and contains no dairy, yeast, corn, or wheat or gluten. Does not contain animal by-products.

#### **PHARMACOLOGICAL ACTIVITIES - TOXICOLOGY - DRUG INTERACTIONS**



# **Cholecalciferol** [Vitamin $D_3$ ] ( $C_{27}H_{44}O$ )

Dogs and cats may have limited ability to use UVB light for cutaneous biogenesis of vitamin D (Kleiman *et al.*, 2010). For this reason, it is important that vitamin  $D_3$  be introduced directly into the diet (Kleiman *et al.*, 2010; Schenck, 2010). Vitamin  $D_3$  regulates calcium:phosphorous balance in the body and it stimulates the kidney's retention of calcium, which is vitally important to bone formation, nerve and muscle control. Osteomalacia occurs when insufficient calcium and/or phosphorus is available for mineralization of newly formed osteoid (Ettinger & Feldman, 2000b).

TOXICOLOGY

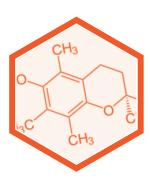
Several factors, such as the chemical form (vitamin  $D_2$  or Vitamin  $D_3$ ), species, dietary intake of calcium and phosphorus, route of administration, and duration of treatment, can influence the maximum tolerable levels of vitamin D in the diet. Most animal species appear to be able to tolerate 10 times the level of vitamin D that they require for long periods of time (NRC, 1987). Intraperitoneal  $LD_{50}$  of vitamin  $D_3$  is 135.4 mg/kg of body weight in male mice (Hatch & Laflamme, 1989). Published human cases of vitamin  $D_3$  toxicity, for which serum levels and dose are known, all involve intake of > or = 40,000 IU (1,000 mcg) per day (Vieth, 1999).

DRUG INTERACTIONS

Due to a narrow therapeutic index, vitamin D analogs given in combination with each other or with pharmacologic doses of vitamin D (calcitriol, doxercalciferol, paricalcitol) may demonstrate additive effects resulting in toxicity manifested as hypercalcemia, hypercalciuria, and hyperphosphatemia (Drugs.com). Atorvastatin appears to increase 25-hydroxycholecalciferol (25[OH]D) concentrations, whereas concurrent vitamin D supplementation decreases concentrations of atorvastatin. Use of thiazide diuretics in combination with calcium and vitamin D supplements may cause hypercalcemia in humans or those with compromised renal function or hyperparathyroidism. Insufficient evidence is available to determine whether lipase inhibitors, antimicrobial agents, antiepileptic drugs, highly active antiretroviral agents, or H2 receptor antagonists alter serum 25(OH)D concentrations (Robien *et al.*, 2013).

# alpha-Tocopherol [Vitamin E] (C<sub>29</sub>H<sub>50</sub>O<sub>2</sub>)

Vitamin E is a natural antioxidant that is important for maintaining stability of cell membranes. Vitamin E supplementation increases antioxidant capacity and can play a potential beneficial role in the prevention or treatment of several diseases in dogs (Raila *et al.*, 2011). Vitamin E deficiency may cause pathologic change in smooth muscle, central nervous system, skeletal muscle, and retina in dogs (Ettinger & Feldman, 2000b). Pansteatitis is associated with a vitamin E deficiency in cats that are habitually or exclusively fed high fat diets, particularly red tuna or other oily fish. Vitamin E supplementation is beneficial in the treatment of pansteatitis (Ettinger & Feldman, 2000a).



**LOXICOLOG** 

Toxicity for vitamin E has not been documented in dogs and cats when administered orally in therapeutic doses. Vitamin E is generally regarded as one of the least toxic fat-soluble vitamins. The oral  $LD_{50}$  alpha-tocopherol acetate for rats, mice and rabbits has been estimated to be >2 g/kg of body weight (NRC, 1987).

## DRUG INTERACTIONS

Vitamin E might slow blood clotting. Taking vitamin E along with anticoagulant drugs, such as warfarin; antiplatelet drugs, such as clopidogrel and dipyridamole; and non-steroidal anti-inflammatory drugs (NSAIDs), including aspirin, ibuprofen, and others might increase the chances of bruising and bleeding. Also, individuals on anticoagulant therapy or individuals who are vitamin K deficient should not take alpha-tocopherol supplements without close medical supervision because of the increased risk of hemorrhage (DRI, 2000). Vitamin E increases xenobiotic pathways that increase hepatic metabolism and excretion of all vitamin K forms (Traber, 2008). A number of medications may decrease the absorption of vitamin E, including cholestyramine, colestipol, isoniazid, mineral oil, orlistat, sucralfate, and the fat substitute, olestra. Anticonvulsant drugs, such as phenobarbital, phenytoin, or carbamazepine, may decrease plasma levels of vitamin E (DRI 2000; Hendler & Rorvik, 2001).

# PHARMACOLOGICAL ACTIVITIES - TOXICOLOGY - DRUG INTERACTIONS



# **beta-Carotene** $(C_{40}H_{56})$

Many animals convert  $\beta$ -carotene to retinol to meet their vitamin A requirements. However, this pathway is inefficient in many carnivores including cats (Schweigert *et al.*, 2002). Although it has been shown that cats are capable of converting  $\beta$ -carotene to active vitamin A, it is inadequate to meet a cat's vitamin A requirement (Green *et al.*, 2011). Dietary supplements of vitamins E and C and beta-carotene reduce oxidative stress in cats with renal insufficiency (Yu & Paetau-Robinson, 2006). Dietary  $\beta$ -carotene stimulates cell-mediated and humoral immune responses in dogs (Chew *et al.*, 2000) and was found to restore immune responses in older dogs (Massimino *et al.*, 2003).

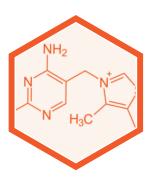
Toxicity for  $\beta$ -carotene has not been documented in dogs and cats when administered orally in therapeutic doses.

#### DRUG INTERACTIONS

Validated interactions studies do not exist for  $\beta$ -carotene preparations. However,  $\beta$ -carotene can interact with medication used for lowering cholesterol. Taking them together can lower the effectiveness of these medications and is considered only a moderate interaction (Web MD, 2012). Orlistat can reduce the absorption of  $\beta$ -carotene by as much as 30% (UMMC, 2012a). Bile acid sequestrants such as cholestyramine and colestipol and proton-pump inhibitors such as omeprazole can also decrease absorption of  $\beta$ -carotene (Meschino, 2012).

# **Thiamine Hydrochloride** [Vitamin B<sub>1</sub>] (C<sub>12</sub>H<sub>17</sub>ClN<sub>4</sub>OS.HCl)

Thiamine (vitamin  $B_1$ ) is an essential cofactor in the decarboxylation of pyruvate and alphaketoglutarate and these reactions are essential for aerobic metabolism. Deficiency of thiamine blocks CNS aerobic metabolic pathways. In cats, initial thiamine deficiency can develop into central vestibular disease, head tremor, mydriasis, and cervical ventroflexion, which may progress to opisthotonos, coma, and death. In dogs, ataxia, paresis, vestibular signs, and seizures have been observed (Ettinger & Feldman, 2000a).



Cats are more susceptible to thiamine deficiency than dogs as they require about four times as much thiamine in the diet. Fish-based diets that contain active thiaminases before processing can destroy thiamine added to these diets and in the heat processing of dog and cat foods, large losses of thiamine can occur. Canned foods often contain gelling agents that increase the pH of the food and in combination with prolonged heat during retorting results in extensive inactivation of thiamine. Moreover, sulphites used for food preservation, cleave the thiamine molecule at the methylene bridge making thiamine inactive. Thiamine deficiency associated with the feeding of meat preserved with sulphur dioxide has been reported in cats and dogs (NRC, 2006).

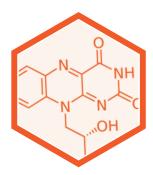
TOXICOLOGY

Toxicity for vitamin  $B_1$  has not been documented in dogs and cats when administered orally in therapeutic doses. Intravenous  $LD_{50}$  of vitamin  $B_1$  is 50-125 mg/kg of body weight in dogs (NRC, 1987).

#### DRUG INTERACTIONS

Validated interactions studies do not exist for thiamine preparations. However, laboratory studies suggest that digoxin may reduce the ability of heart cells to absorb and utilize thiamine. Diuretics such as furosemide may reduce the levels of thiamine in the body (UMMC, 2012b).

#### PHARMACOLOGICAL ACTIVITIES - TOXICOLOGY - DRUG INTERACTIONS



# **Riboflavin** [Vitamin $B_2$ ] ( $C_{17}H_{20}N_4O_6$ )

The major function of riboflavin is to serve as a precursor of the coenzymes flavin mononucleotide (FMN) and flavin adenine dinucleotide (FAD). Synthesis of the coenzymes from riboflavin is under the control of thyroid hormones. A deficiency of riboflavin impacts other vitamins because flavin coenzymes are involved in their metabolism. These vitamins include folic acid, pyridoxine, niacin, and vitamins K and D. Acute riboflavin deficiency in dogs and cats results in anorexia, body weight loss, decreased activity, hypothermia, decreased respiratory rate, progressive weakness, ataxia, sudden collapse to semicomatose state, and death. Chronic riboflavin deficiency has been associated with anorexia, body weight loss, muscular weakness, flaking dermatitis of the abdomen and medial surface of the hind legs, and ocular lesions (NRC, 2006).

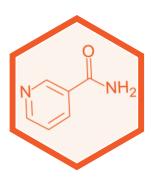
Toxicity for vitamin  $B_2$  has not been documented in dogs and cats when administered orally in therapeutic doses. Oral  $LD_{50}$  of vitamin  $B_2$  is >10 g/kg of body weight in rats (NRC, 1987).

#### DRUG INTERACTIONS

Validated interactions studies do not exist for riboflavin preparations. However, anticholinergic drugs and probenecid may decrease riboflavin absorption. Riboflavin can affect the absorption of tetracycline. Tricyclic antidepressants, phenothiazines, doxorubicin, and phenytoin may reduce levels of riboflavin in the body. Methotrexate can inhibit the utilization of riboflavin (UMMC, 2012c).

# **Niacinamide** [Vitamin $B_3$ ] ( $C_6H_6N_2O$ )

Dogs and cats derive most of their energy from oxidation-reduction (redox) reactions, which are processes involving the transfer of electrons. As many as 200 enzymes require the niacin coenzymes, nicotinamide adenine dinucleotide (NAD) and nicotinamide adenine dinucleotide phosphate (NADP), mainly to accept or donate electrons for redox reactions. NAD functions most often in energy producing reactions involving the catabolism of carbohydrates, fats, proteins, and alcohol. NADP functions more often in anabolic reactions, such as in the synthesis of all macromolecules, including fatty acids and cholesterol (Brody, 1999).



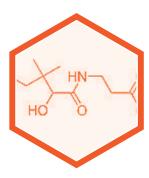
A vitamin B<sub>3</sub> deficiency is characterized by loss of appetite, fatigue, apathy and mouth ulcers, diarrhoea, emaciation and pruritic dermatitis of the hind legs and ventral abdomen. Cats need more niacin than dogs. Dogs are able to synthesize nicotinamide endogenously from tryptophan; cats do not produce any measurable quantities. Cats possess all the enzymes of the pathway of niacin synthesis, but the activity of picolinic carboxylase is extremely high, precluding any measurable synthesis of nicotinic acid (NRC, 2006; Ettinger & Feldman, 2000a).

тохісогобу

Toxicity for vitamin  $B_3$  has not been documented in dogs and cats when administered orally in therapeutic doses. However, it has been reported that repeated oral administration of 2 g/day of nicotinic acid produced bloody feces, convulsions and death in a few dogs (Chen *et al.*, 1938). Oral LD<sub>50</sub> of niacinamide is 4.5-7 g/kg of body weight in rats (NRC, 1987).

# DRUG INTERACTIONS

Validated interactions studies do not exist for niacinamide preparations. However, niacinamide inhibits metabolism of primidone in mice and metabolism of primidone and carbamazepine in humans. This probably occurs by inhibition of cytochrome P-450 by nicinamide (Bourgeois *et al.*, 1982).



# **Calcium Pantothenate** [Vitamin $B_5$ ] ( $C_{18}H_{32}CaN_2O_{10}$ )

Pantothenic acid is found throughout living cells in the form of coenzyme A (CoA), a vital coenzyme in numerous chemical reactions (Tahiliani & Beinlich, 1991). CoA is required for chemical reactions that generate energy from fat, carbohydrates, and proteins. The synthesis of essential fats, cholesterol, and steroid hormones requires CoA, as does the synthesis of melatonin and the neurotransmitter, acetylcholine. Heme requires succinyl-CoA for its synthesis and the metabolism of a number of drugs and toxins by the liver requires CoA (Brody, 1999). Failure to grow and histological changes are the main signs of pantothenate deficiency in cats (NRC, 2006).

Toxicity for vitamin  $B_5$  has not been documented in dogs and cats when administered orally in therapeutic doses. Oral  $LD_{50}$  of calcium pantothenate is 10 g/kg of body weight in mice (Unna & Greslin, 1941)

**DRUG** Validated interactions studies do not exist for calcium pantothenate preparations. Clinical interactions **INTERACTIONS** with other drugs have not been reported.

# **Pyridoxine Hydrochloride** [Vitamin B<sub>6</sub>] (C<sub>8</sub>H<sub>11</sub>NO<sub>3</sub>·HCl)

The liver is the primary organ for metabolism of vitamin  $B_6$ , and it releases the active form pyridoxal-5-phosphate (PLP) into the circulation to supply other tissues. Low riboflavin status leads to a reduction in circulating PLP (NRC, 2006). PLP plays a vital role in the function of approximately 100 enzymes that catalyze essential chemical reactions in the body. PLP functions as a coenzyme for glycogen phosphorylase, an enzyme that catalyzes the release of glucose from stored glycogen. Much of the PLP in the body is found in muscle bound to glycogen phosphorylase. PLP is also a coenzyme for reactions used to generate glucose from amino acids, a process known as gluconeogenesis (Shils *et al.*, 2006).



Vitamin  $B_6$  deficiency can result in growth depression, a mild microcytic hypochromic anemia with elevated serum iron; convulsive seizures, and irreversible kidney lesions consisting of tubular atrophy and dilation, fibrosis, and intratubular deposits of birefringent crystals (NRC, 1986). It may also cause a dull, waxy, unkempt coat with fine scales and patchy alopecia (Watson, 1998).

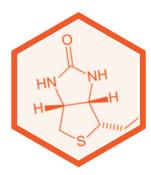
TOXICOLOGY

Toxicity for vitamin  $B_6$  has not been documented in dogs and cats when administered orally in therapeutic doses. However, studies have suggested that probably 1,000 times the nutritional requirements would have to be included in diets in order to produce signs of toxicity. The LD<sub>50</sub> for vitamin  $B_6$  in dogs is 1 g/kg of body weight. According to another report, dogs can tolerate up to 1 g of pyridoxine/kg of body weight for a short duration of time, but over longer periods can cause ataxia, muscle weakness and neurological damage (Friedrich, 1988). In another study dogs administered 50 mg of pyridoxine hydrochloride/kg of body weight reported no signs of toxicity (Phillips *et al.*, 1978). Levels of pyridoxine of 1,000 mg/kg of diet for less than 60 days, or less than 500 mg/kg of diet for more than 60 days, appear to be safe for dogs (NRC, 1987).

## DRUG INTERACTIONS

Pyridoxine deficiency can occur with medications such as isoniazid, cycloserine, and penicillamine. High doses of pyridoxine may decrease the efficacy of phenobarbital and phenytoin (Shils *et al.*, 2006; Bender, 1999). Pyridoxine 10-25 mg may be enough to inhibit levodopa (Leon *et al.*, 1971).

## PHARMACOLOGICAL ACTIVITIES - TOXICOLOGY - DRUG INTERACTIONS



# **Biotin** [Vitamin $B_7$ ] ( $C_{10}H_{16}N_2O_3S$ )

Biotin is attached at the active site of five mammalian enzymes known as carboxylases. Acetyl-CoA carboxylase I and II catalyze the binding of bicarbonate to acetyl-CoA to form malonyl-CoA which is required for the synthesis of fatty acids. Pyruvate carboxylase is a critical enzyme in gluconeogenesis—the formation of glucose from sources other than carbohydrates. Methylcrotonyl-CoA carboxylase catalyzes an essential step in the catabolism of leucine, an essential amino acid. Propionyl-CoA carboxylase catalyzes essential steps in the metabolism of certain amino acids, cholesterol, and odd chain fatty acids (Chapman-Smith & Cronan, 1999; Zempleni & Mock, 1999). Symptoms of biotin deficiency include dried secretions around the eyes, nose and the angle of the mouth, scaly dermatitis, alopecia, hypersalivation, bloody diarrhea, anorexia, and emaciation (NRC, 1986).

TOXICOLOG

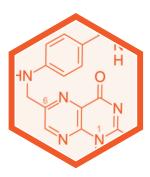
Toxicity for biotin has not been documented in dogs and cats when administered orally in therapeutic doses. The  $LD_{50}$  for repeated (10 day) oral administration in rats was found to be >350 mg/day (EVM, 2002).

# DRUG INTERACTIONS

Anticonvulsant drugs such as primidone and carbamazepine inhibit biotin absorption in the small intestine. Use of valproic acid has been associated with decreased biotinidase activity (Schulpis *et al.*, 2001; Bowman & Russel, 2006).

# Folic Acid [Vitamin $B_9$ ] ( $C_{19}H_{19}N_7O_6$ )

The terms folic acid and folate are often used interchangeably, folic acid, the more stable form, occurs rarely in foods. The function of folate coenzymes in the body is to mediate the transfer of one-carbon units (Choi & Mason, 2000). Folate coenzymes act as acceptors and donors of one-carbon units in a variety of reactions critical to the metabolism of nucleic acids and amino acids (Bailey & Gregory, 1999). Deficiency of folic acid can cause macrocytic anemia, megaloblastic anemia, leucopenia, elevated plasma iron concentration, and weight loss (NRC, 1986).

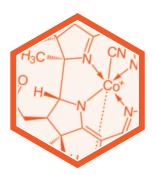


TOXICOLOGY

Toxicity for folic acid has not been documented in dogs and cats when administered orally in therapeutic doses. The Registry of toxic effects of chemical substances (US Department of Health and Human Services, 1979) gives two  $LD_{50}$  values for folic acid in mice, intraperitoneal 100 mg/kg of body weight and intravenous 239 mg/kg of body weight (Parchure *et al.*, 1985).

#### DRUG INTERACTIONS

NSAIDs such as aspirin or ibuprofen may interfere with folate metabolism when taken in very large therapeutic dosages. Phenytoin, has been shown to inhibit the intestinal absorption of folate, and several studies have associated decreased folate status with long-term use of the anticonvulsants, phenytoin, phenobarbital, and primidone (Lewis *et al.*, 1995; Apeland *et al.*, 2001). Taking folic acid at the same time as the cholesterol-lowering agents such as cholestyramine and colestipol, may decrease the absorption of folic acid (Hendler & Rorvik, 2001). A number of other medications have been shown to have antifolate activity, including tetracycline, trimethoprim, pyrimethamine, triamterene, and sulfasalazine (UMMC, 2012d).



# **Cyanocobalamin** [Vitamin $B_{12}$ ] ( $C_{63}H_{88}CoN_{14}O_{14}P$ )

Vitamin  $B_{12}$  has the largest and most complex chemical structure of all the vitamins. It is unique among vitamins in that it contains a metal ion, cobalt. In mammals, cobalamin is a cofactor for only two enzymes, methionine synthase and L-methylmalonyl-CoA mutase (Shils *et al.*, 2006). A deficiency of cobalamin can cause a clinical and hematologic picture identical to that of folate deficiency (Ettinger & Feldman, 2000b). There are a number of clinical reports of cobalamin deficiency in dogs and these reports relate to either deficiencies induced by bacterial overgrowth of the intestine resulting in decreased availability of cobalamin or genetic abnormalities of cobalamin metabolism (NRC, 2006).

ΤΟΧΙΟΟΙΟΘΥ

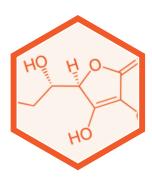
Toxicity for vitamin  $B_{12}$  has not been documented in dogs and cats when administered orally in therapeutic doses. However, subcutaneous doses of 2 to 33 µg/kg of body weight have been given to dogs, and have resulted in disturbances of reflex activity (NRC, 2006). Oral LD<sub>50</sub> for cyanocobalamin is 13,500 mg/kg of body weight in rats and 22,000 mg/kg of body weight in mice (USP, 2007).

## DRUG INTERACTIONS

Medications that reduce levels of vitamin  $B_{12}$  in the body include proton pump inhibitors such as esomeprazole, lansprazole, omeprazole and rabeprazole (Kasper, 1999);  $H_2$  blockers including cimetidine, famotidine, and ranitidine (Termanini *et al.*, 1998); anti-seizure medications such as phenytoin, phenobarbital, and primidone; anti-diabetic medication metformin; bile acid sequestrants including colestipol, cholestyramine, and colsevelam (UMMC, 2012e).

# Ascorbic Acid [Vitamin C] (C<sub>6</sub>H<sub>8</sub>O<sub>6</sub>)

Vitamin C is required for the synthesis of collagen, an important structural component of blood vessels, tendons, ligaments, and bone. Vitamin C also plays an important role in the synthesis of the neurotransmitter, norepinephrine. Vitamin C deficiency can slow healing and increase susceptibility to disease. In dogs and cats supplementation of vitamin C may be beneficial, particularly in times of stress. Studies have shown a slight depression in serum vitamin C concentration in response to some conditions and diseases, and supplementation can increase serum levels in healthy animals (NRC, 2006).



**LOXICOLOGY** 

Toxicity for vitamin C has not been documented in dogs and cats when administered orally in therapeutic doses. Oral  $LD_{50}$  for ascorbic acid is 11,900 mg/kg in rat and 3,367 mg/kg of body weight in mice (ACROS, 2008).

# DRUG INTERACTIONS

Estrogen-containing contraceptives and aspirin can lower vitamin C levels in plasma and white blood cells (Basu, 1982). High doses of vitamin C have been found to interfere with the interpretation of certain laboratory tests such as serum bilirubin, serum creatinine, and the guaiac assay for occult blood (Hendler & Rorvik, 2001).



## Calcium (Citrate)

Calcium is a major structural element in bones and teeth. The amount of calcium absorption in dogs ranges from 25 to 90 percent, depending on the amount of intake and the age of the animal (Ettinger & Feldman, 2000a). Calcium deficiency in dogs is characterized by rickets in normal dogs, milk fever syndrome in pregnant or lactating dogs and a condition known as nutritional secondary hyperparathyroidism (NSHP). Chronic dietary calcium deficiency causes major decreases in bone material content, which can result in significant skeletal abnormalities including fractures. Calcium intake is tied directly to the calcium-phosphorus ratio (1.5:1) in the body. A diet high in calcium and low in phosphorus may lead to problems metabolizing the calcium. It will cause bone deformities and hip dysplasia. Calcium deficiency in kittens demonstrated bone rarefaction, especially in the lumbar vertebrae which tended to curve and collapse, and in the pelvis (NRC, 2006).

# TOXICOLOG

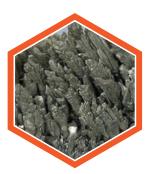
Toxicity for calcium citrate has not been documented in dogs and cats when administered orally in therapeutic doses.  $LD_{50}$  of calcium citrate is not documented. Oral  $LD_{50}$  for calcium carbonate is 6,450 mg/kg of body weight in rats (Sciencelab, 2010a).

#### DRUG INTERACTIONS

Significant interactions have been observed between calcium and certain antibiotics namely tetracyclines and fluoroquinolones (Pfizer, 1990; Bayer, 2002). Calcium decreases the bioavailability of levothyroxine (Abbott, 2002). Combining calcium with thiazide diuretics increases the risk of developing hypercalcemia. High doses of supplemental calcium could increase the likelihood of abnormal heart rhythms in people taking digitalis for heart failure (Vella *et al.*, 1999). Intravenous calcium salts can prevent hypotension associated with intravenous verapamil (Moser *et al.*, 2000). Calcium citrate when taken with aluminum-containing antacids, the amount of aluminum absorbed into the blood may be increased significantly. Bile acid sequestrants such as cholestyramine, colestipol, and colesevelam may interfere with calcium absorption and increase the loss of calcium in the urine (UMMC, 2012f).

# Magnesium (Citrate)

Magnesium is involved in more than 300 essential metabolic reactions such as energy production, synthesis of essential molecules (nucleic acid, enzymes, and glutathione), structural roles (cell membranes, chromosomes), and ion transport across cell membranes, cell signalling, and cell migration (Shils *et al.*, 2006). Magnesium deficiency in dogs can cause anorexia, weight loss, hyperextension of the carpal joints and hind-leg paralysis. Magnesium deficiency in cats can cause poor growth rate and overextension of the metacarpi followed by muscular twitching and convulsions (NRC, 2006).



ΓΟΧΙΟΟΙΟΘΥ

Toxicity for magnesium citrate has not been documented in dogs and cats when administered orally in therapeutic doses.  $LD_{50}$  for magnesium sulphate in dogs is considered to be >1,200 mg/kg of body weight [infused at 200 mg/kg/hr] (Mochizuki *et al.*, 1998).

#### DRUG INTERACTIONS

The absorption of quinolone antibiotics, such as ciprofloxacin and moxifloxacin, tetracycline antibiotics, including tetracycline, doxycycline, and minocycline, and nitrofurantoin, may be diminished when taking magnesium supplements. Magnesium may potentiate the side effects such as dizziness, nausea, and fluid retention associated with calcium channel blockers. High doses of furosemide and some thiazide diuretics, if taken for extended periods, may result in magnesium depletion (Hendler & Rorvik, 2001; UMMC, 2012). Digoxin can lead to increased loss of magnesium in the urine and it is important that normal levels of magnesium be maintained while taking digoxin because low blood levels of magnesium can increase adverse effects from this drug, including heart palpitations and nausea (UMMC, 2012g).

## PHARMACOLOGICAL ACTIVITIES - TOXICOLOGY - DRUG INTERACTIONS



# **Boron** (Sodium Borate)

Boron acts directly or indirectly to influence the composition, structure and strength of bones. Experiments in animals indicate that boron may influence calcium, phosphorus, magnesium, and cholecalciferol metabolism. A vitamin D deficiency enhances the need for boron, and boron normalizes abnormalities associated with a magnesium deficiency. Boron also has an effect on brain electrical activity (Groff *et al.*, 1995; NRC, 2006). The signs of boron deficiency in animals include depressed growth and in humans, a low-boron diet is associated with increased urinary calcium and magnesium excretion and with alteration in steroid hormone metabolism (Groff *et al.*, 1995).

Toxicity for boron has not been documented in dogs and cats when administered orally in therapeutic doses. A study found that daily doses of 3 g of boric acid, 5 g of borax for 8-10 days had no physiological or pharmacological effect in adult dogs weighing 8 to 12 kg (NRC, 2005). Oral  $LD_{50}$  for sodium borate is 4.5 g/kg of body weight and 4.98 g/kg of body weight in male and female rats respectively (Weir & Fisher, 1999).

**DRUG** Validated interactions studies do not exist for boron preparations. Clinical interactions with other **INTERACTIONS** drugs have not been reported.

# Chromium (Polynicotinate)

Chromium is an essential dietary trace mineral involved in carbohydrate and lipid metabolism. Chromium is required for cellular uptake of glucose, and chromium deficiency causes insulin resistance. Chromium supplementation may improve insulin sensitivity and has been used as adjunct treatment of diabetes mellitus in humans. Chromium supplementation in diabetic dogs at a level of 2.5-7.5  $\mu$ g/kg for several months was found to be safe (Schachter *et al.*, 2001). Chromium has also been shown to be involved in protein synthesis, and nucleic acid metabolism (NRC, 2006). Signs of chromium deficiency in humans and several animal species have been found to include impaired glucose tolerance, elevated plasma insulin, hyperglycemia, impaired growth, elevated plasma triglycerides, neuropathy, encephalopathy, corneal lesions and decreased fertility and sperm count.



TOXICOLOGY

Toxicity for chromium polynicotinate has not been documented in dogs and cats when administered orally in therapeutic doses. Hexavalent chromium or chromium ( $Cr^{+6}$ ) is a recognized carcinogen. In contrast, there is little evidence that trivalent chromium ( $Cr^{+3}$ ) is toxic to humans (FNB & IOB, 2001b). Intravenous  $LD_{50}$  for nicotinic acid bound chromium complexes is approximately 1 g/kg of body weight in rats (EFSA, 2008).

# DRUG INTERACTIONS

Validated interactions studies do not exist for chromium preparations. Clinical interactions with other drugs have not been reported. However, large doses of calcium carbonate or magnesium hydroxide-containing antacids decreased chromium absorption in rats. In contrast, aspirin and indomethacin both increased chromium absorption in rats (Shils *et al.*, 2006).



# Copper (Gluconate)

Copper is an essential trace element for humans and animals. In the body, copper shifts between the cuprous (Cu<sup>1+</sup>) and cupric (Cu<sup>2+</sup>) forms, though the majority of the body's copper is in the Cu<sup>2+</sup> form. Copper is a critical functional component of a number of essential enzymes known as cuproenzymes. Some of the physiologic functions known to be copper-dependent include energy production, regulation of gene expression (Uauy *et al.*, 1998), formation of connective tissue, the pigment melanin and maintenance of myelin sheath (Shils *et al.*, 2006), iron metabolism, synthesis of the neurotransmitter norepinephrine (O'Dell & Sunde, 1997), metabolism of neurotransmitters norepinephrine, epinephrine, dopamine and serotonin (FNB & IOB, 2001), antioxidant functions such as copper dependent superoxide dismutase and ceruloplasmin (Johnson *et al.*, 1992). In dogs copper deficiency can cause lameness and bone fragility, loss of hair pigmentation, and hyperextension in the distal phalanges (Ettinger & Feldman, 2000b; NRC, 2006).

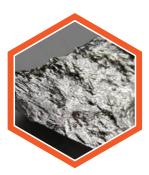
Toxicity for copper gluconate has not been documented in dogs and cats when administered orally in therapeutic doses. However, in various breeds of dogs, especially Bedlington Terriers, an inherited sensitivity to copper toxicosis similar to Wilson's disease in humans has been identified. Acute poisoning is usually seen after accidental administration of excessive amounts of soluble copper salts (Kahn & Line, 2010). Intradermal  $LD_{50}$  for copper salts is >1124 mg/kg of body weight in rats (EVM, 2000).

## DRUG INTERACTIONS

Validated interactions studies do not exist for copper preparations. However, penicillamine dramatically increases the urinary excretion of copper; individuals taking the medication for reasons other than copper overload may have an increased copper requirement. Additionally, antacids may interfere with copper absorption when used in very high amounts (Shils et al., 2006).

# Manganese (Sulphate)

Manganese plays an important role in a number of physiologic processes as a constituent of metalloenzymes and as an enzyme activator (Groff *et al.*, 1995). The functions of manganese include metabolism of carbohydrates, amino acids and cholesterol (FNB & IOB, 2001a), a component of manganese superoxide dismutase which is the principal antioxidant enzyme in the mitochondria (O'Dell & Sunde, 1997), formation of cartilage and bone (Ziegler & Filer, 1996; Ettinger & Feldman, 2000a), synthesis of glycosaminoglycan and production of collagen required in wound healing (Klimis-Tavantzis, 1994; Muszyńska *et al.*, 2000). Manganese deficiency is very rare in dogs and cats. Experimental dietary deficiency leads to disproportionate, shortened, and thickened long bones in different species; defective development of the skull; and formation of otoliths in the inner ear during gestation (Ettinger & Feldman, 2000a).



**JOXICOLOG** 

Toxicity for manganese sulphate has not been documented in dogs and cats when administered orally in therapeutic doses. However, in a study intravenous infusion of manganese chloride 16 mg/kg/day (3-5 times daily dose of manganese) to beagle dogs caused severe hepatotoxicity (Khan *et al.*, 1997). Oral  $LD_{50}$  for manganese sulphate is 2,150 mg/kg of body weight in rats (MDL, 2002).

# DRUG INTERACTIONS

Validated interactions studies do not exist for manganese preparations. However, magnesium-containing antacids and laxatives and the antibiotic medication, tetracycline, may decrease the absorption of manganese if taken together with manganese-containing foods or supplements (Hendler & Rorvik, 2001).



# **Selenium** (L-Selenomethionine)

Humans and animals require selenium for the function of a number of selenium-dependent enzymes, also known as selenoproteins. During selenoprotein synthesis, selenocysteine is incorporated into a very specific location in the amino acid sequence in order to form a functional protein. At least 25 selenoproteins have been identified, but the metabolic functions have been identified for only about one-half of them. The selenoproteins with an identified function include: glutathione peroxidises which are antioxidant enzymes (Gladyshev, 2006); thioredoxin reductase which participates in the regeneration of several antioxidants including vitamin C (Mustacich & Powis, 2000); iodothyronine deiodinases involved in the regulation of thyroid hormones (Hatfield et al., 2006); selenoprotein P, associated with protection of vascular endothelial cells against reactive nitrogen species (Arteel et al., 1999); selenoprotein W, thought to play a role in muscle growth and differentiation by protecting the developing myoblast from oxidative stress (Loflin et al. 2006); selenoprotein V functions in spermatogenesis; selenoprotein S is involved with inflammatory and immune responses; kDA selenoprotein has a redox function and is implicated in cancer prevention (Gladyshev, 2006). Animal studies indicate that selenium and vitamin E tend to spare one another and that selenium can prevent some of the damage resulting from vitamin E deficiency in models of oxidative stress (Sword et al., 1991). Only one paper reports experimentally produced clinical signs of selenium deficiency in dogs; there are no reports for cats. Clinical signs include anorexia, depression, dyspnea, and coma (NRC, 2006).

ΤΟΧΙΟΟΙΟΟΥ

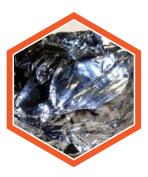
Toxicity for selenium has not been documented in dogs and cats when administered orally in therapeutic doses. However, a single acute oral dose of selenium in the range of 1-5 mg/kg of body weight is lethal in most animals. Parenteral selenium products are also quite toxic, especially to young animals, and have caused deaths in baby pigs, calves, and dogs at doses as low as 1.0 mg/kg of body weight (Kahn & Line, 2010). Oral LD<sub>50</sub> for sodium selenite is 1.0 mg/kg in rabbits, 3.0 mg/kg of body weight in mice and 4.8-7.0 mg/kg of body weight in rats (EMEA, 1997). Vitamin E exhibits a protective effect on selenium intoxication (Berschneider *et al.*, 1976).

#### DRUG INTERACTIONS

Validated interactions studies do not exist for selenium preparations. However, the anticonvulsant medication valproic acid has been found to decrease plasma selenium levels. Animal studies have found that supplemental sodium selenite decreases the toxicities of the antibiotic nitrofurantoin (Flodin, 1990).

# Silicon (Sodium Metasilicate)

Silicon is an essential nutrient that plays a role in the calcification and maturation of bone. Silicon also appears to be a cofactor in prolyl hydrase activity, which is involved in collagen synthesis. Signs of silicon deficiency are related mainly to aberrant development of connective tissue and bone (NRC, 2006).



TOXICOLOGY

Toxicity for silicon has not been documented in dogs and cats when administered orally in therapeutic doses. No significant acute toxicity or mortality has been reported in animals given doses up to 3,000 mg/kg of body weight per day (NRC, 2005). Oral  $LD_{50}$  for sodium metasilicate is 1,280 mg/kg of body weight in rats and 2,400 mg/kg in mice (Haneke, 2002).

**DRUG** Validated interactions studies do not exist for silicon preparations. Clinical interactions with other drugs have not been reported.



# Zinc (Citrate)

Zinc plays important roles in growth and development, the immune response, neurological function, and reproduction. On the cellular level, the function of zinc can be divided into three categories: catalytic, structural, and regulatory (Bowman & Russell, 2006). Inadequate zinc supply, especially in growing animals, may lead to severe clinical signs within days, resulting in growth depression, skin defects, impaired immune function, and growth disorders of the skeleton (Ettinger & Feldman, 2000a). Zinc deficiency in the dog most commonly occurs as a skin condition called 'zinc responsive dermatosis' (Colombini, 1999; Campbell & Crow, 2010). The usual symptoms are hair loss, and scaling and crusting of the skin around the face, head, and legs. Lesions often encircle the mouth, chin, eyes, and ears. The foot pads may be scaly and the hair coats are dull and dry.

Toxicity for zinc has not been documented in dogs and cats when administered orally in therapeutic doses. However, zinc toxicosis caused by ingestion of foreign materials such as galvanized metal and pennies has been reported in dogs, but it has not been described in cats (Hardy *et al.*, 2003). No evidence of significant pathological effects were observed in rats following daily oral administration of zinc (as zinc citrate, zinc acetate, zinc oxide or zinc maleate) in doses up to 34 mg/kg of body weight for 35 to 53 weeks. Oral LD<sub>50</sub> for zinc chloride is 350 mg/kg of body weight in rats and mice, 200-250 mg/kg of body weight in rabbits. Oral LD<sub>50</sub> for zinc sulphate is 2,200 mg/kg of body weight in rats and 2,100 mg/kg of body weight in rabbits (EMEA, 1996).

#### DRUG INTERACTIONS

Both quinolone antibiotics and tetracycline antibiotics interact with zinc in the gastrointestinal tract, inhibiting the absorption of both zinc and the antibiotic (Lomaestro *et al.*, 1995; Penttilä *et al.*, 1975). Zinc can reduce the absorption and action of penicillamine, a drug used to treat rheumatoid arthritis (Brewer *et al.* 1993). Thiazide diuretics such as chlorthalidone and hydrochlorothiazide increase urinary zinc excretion by as much as 60% (Wester, 1980).

# **Choline** [Bitartrate] $(C_9H_{19}NO_7)$

Although choline is not by strict definition a vitamin, it is an essential nutrient (Brody, 1999). The initial recognition of choline as a significant dietary factor depended on its capacity to reduce the fat content of the liver of diabetic dogs (Goodman & Gilman, 1996). The majority of the body's choline is found in specialized fat molecules known as phospholipids. Choline and its metabolites serve a number of vital biological functions such as structural integrity of cell membranes, cell signalling, nerve impulse transmission, lipid transport and metabolism (Groff *et al.*, 1995). Choline deficiency in animals can result in weight gain, fatty liver, liver cirrhosis and in severe cases, vomiting and death. Other effects of a choline deficiency include reduced plasma choline concentration, thymus atrophy, decreased growth rate, impaired reproduction, and hemorrhagic renal lesions. A chronic deficiency in dogs has been reported to cause anaemia, duodenal ulcers, liver damage and oedema (Lewis, 2005).

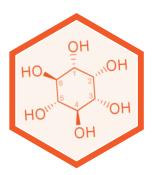


TOXICOLOGY

Toxicity for choline has not been documented in dogs and cats when administered orally in therapeutic doses. However, adverse effects have been reported for levels of choline chloride equivalent to 3 times the choline requirement (NRC, 1987).  $LD_{50}$  for choline bitartrate has not been documented. Oral  $LD_{50}$  for choline chloride is 3,900 mg/kg of body weight in mice (NRC, 1987).

# DRUG INTERACTIONS

Validated interactions studies do not exist for choline preparations. Clinical interactions with other drugs have not been reported. However, in animal model, methotrexate administration has shown to deplete choline (Shils *et al.*, 2006).



# **Inositol** $(C_6H_{12}O_6)$

Inositol and its isomers function as the basis for a number of signalling and secondary messenger molecules. They are involved in a number of biological processes, including insulin signal transduction (Larner, 2002), nerve transmission, intracellular calcium concentration control (Gerasimenko *et al.*, 2006), metabolism of fats and reducing blood cholesterol (Rapiejko *et al.*, 1986), cell membrane potential maintenance (Kukuljan *et al.*, 1997), serotonin activity modulation (Einat *et al.*, 2001), and gene expression, (Shen *et al.*, 2003; Steger *et al.*, 2003). No deficiencies of inositol have been documented in either dogs or cats (NRC, 2006).

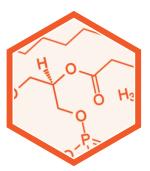
TOXICOLOG

Toxicity for inositol has not been documented in dogs and cats when administered orally in therapeutic doses. Oral  $LD_{50}$  for inositol is 10 g/kg of body weight in mice (Sciencelab, 2010b).

DRUG Validated interactions studies do not exist for inositol preparations. Clinical interactions with otherINTERACTIONS drugs have not been reported.

# Lecithin

Lecithin is a generic term to designate any group of yellow-brownish fatty substances occurring in animal and plant tissues, and in egg yolk, composed of phosphoric acid, choline, fatty acids, glycerol, glycolipids, triglycerides, and phospholipids. Phosphatidylcholine occurs in all cellular organisms, being one of the major components of the phospholipid portion of the cell membrane. Lecithin supplementation decreases hyperlipidemia, influences lipid metabolism, exhibits hepatoprotective (Lamireau *et al.*, 2007) and immunomodulatory (Miranda *et al.*, 2008) effects.



TOXICOLOG

Toxicity for lecithin has not been documented in dogs and cats when administered orally in therapeutic doses. Oral  $LD_{s_0}$  for lecithin is >8 ml/kg of body weight in rats (Pfizer, 2009).

# DRUG INTERACTIONS

Validated interactions studies do not exist for lecithin preparations. Clinical interactions with other drugs have not been reported.

PRECAUTIONS	• An examination from a veterinarian is recommended prior to using this produ		
	• Safe use in pregnant animals or animals intended for breeding has not been proven.		
	• If animal's condition worsens or does not improve, stop product administration and consult your veterinarian.		
	• Not to be used one week prior to surgery.		
	• Consult your veterinarian for potential drug interactions.		
	• Off-label use of this product in ruminants is not recommended.		
	• Oral use only.		
	• Administer during or after the animal has eaten to reduce incidence of gastrointestinal upset.		
	• Shake well before use.		
WARNINGS	<ul> <li>To be used in dogs and cats only.</li> <li>Keep out of reach of children and animals.</li> <li>In case of accidental overdose, contact a health professional immediately.</li> </ul>		
ADVERSE REACTIONS	• Mild gastrointestinal discomfort may occur which is dose dependent.		
CONTRAINDICATIONS	• None documented.		





#### REFERENCES

Abbott Laboratories. (2002). Product Information: Synthroid, levothyroxine. Oral tablet, USP, levothyroxine sodium oral tablet, USP, North Chicago, IL.

Acros Organics. (2008). Material Safety Data Sheet. L(+)-Ascorbic Acid. Accessed: January 8, 2012. Available at: http://www.ch.ntu.edu.tw/~genchem99/msds/exp12/ vitamin%20C.pdf

Apeland T, Mansoor MA, Strandjord RE. (2001). Antiepileptic drugs as independent predictors of plasma total homocysteine levels. Epilepsy Research. 47(1-2):27-35.

Arteel GE, Briviba K, Sies H. (1999). Protection against peroxynitrite. FEBS Letters. 445: 226-30.

Bailey LB, Gregory JF, 3rd. (1999). Folate metabolism and requirements. Journal of Nutrition. 129: 779-782.

Basu TK. (1982). Vitamin C-aspirin interactions. International Journal for Vitamin and Nutrition Research. Supplement. 23:83-90.

Bayer Corporation. (2002). Product Information: Cipro, ciprofloxacin., West Haven, CT.

Bender DA. (1999). Non-nutritional uses of vitamin B6. The British Journal of Nutrition. 81(1):7-20.

Berschneider F, Hess M, Neuffer K, Willer S. (1976). LD<sub>50</sub> values and selenium concentration in rabbit organs after parenteral administration of sodium selenite and determination of toxicity of urososelevit pro inj (in German). Archiv fur Experimentelle Veterinarmedizin. 30:627-32.

Bourgeois BF, Dodson WE, Ferrendelli JA. (1982). Interactions between primidone, carbamazepine, and nicotinamide. Neurology. 32(10):1122-26.

Bowman BA, Russell RM. (Eds). (2006). Present Knowledge in Nutrition. 9th Edition, Vol. 1. Washington, D.C.: ILSI Press.

Brewer GJ, Yuzbasiyan-Gurkan V, Johnson V, Dick RD, Wang Y. (1993). Treatment of Wilson's disease with zinc: XI. Interaction with other anticopper agents. Journal of the American College of Nutrition. 12:26-30.

Brody T. (1999). Nutritional Biochemistry. 2nd ed. San Diego: Academic Press.

Campbell GA, Crow D. (2010). Severe zinc responsive dermatosis in a litter of Pharaoh Hounds. Journal of Veterinary Diagnostic Investigation. 22:663-6.

Campbell NR, Hasinoff BB, Stalts H, et al. (1992). Ferrous sulfate reduces thyroxine efficacy in patients with hypothyroidism. Annals of Internal Medicine. 117:1010-1013.

Chapman-Smith A, Cronan JE, Jr. (1999). Molecular biology of biotin attachment to proteins. Journal of Nutrition. 129:4775-4845.

Chen KK, Rose CL, Robbins EB. (1938). Toxicity of Nicotinic Acid. Experimental Biology and Medicine. 38:241-245.

Chew BP, Park JS, Wong TS, et al. (2000). Dietary beta-carotene stimulates cell-mediated and humoral immune response in dogs. The Journal of Nutrition. 130:1910-3.

Choi SW, Mason JB. (2000). Folate and carcinogenesis: an integrated scheme. Journal of Nutrition. 130:129-132.

Colombini S. (1999). Canine zinc-responsive dermatosis. The Veterinary Clinics of North America. Small Animal Practice. 29:1373-83.

DRI. (2000). Food and Nutrition Board, Institute of Medicine. Vitamin E. Dietary reference intakes for vitamin C, vitamin E, selenium, and carotenoids. Washington D.C.: National Academy Press.

Drugs.com. Accessed: January 9th, 2012. Available at: http://www.drugs.com/drug-interactions/multivitamin-with-minerals-index.html?filter=3&generic\_only=

Einat H, Clenet F, Shaldubina A, *et al.* (2001). The antidepressant activity of inositol in the forced swim test involves 5-HT(2) receptors. Behavioural Brain Research. 118:77-83.

EMEA. (1996). The European Agency for the Evaluation of Medicinal Products. Veterinary Medicines Evaluation Unit. Committee for Veterinary Medicinal Products. Zinc Salts Summary Report. EMEA/MRL/113/96-Final. London, UK.

EMEA. (1997). The European Agency for the Evaluation of Medicinal Products. Veterinary Medicines Evaluation Unit. Committee for Veterinary Medicinal Products. Potassium and Sodium Salts of Selenium. Summary Report. EMEA/MRL/249/97-Final. London, UK.

Ettinger SJ, Feldman EC. (2000a). Textbook of Veterinary Internal Medicine. Diseases of the Dog and Cat. Fifth Edition, Volume 1. Philadelphia: W.B. Saunders Company.

Ettinger SJ, Feldman EC. (2000b). Textbook of Veterinary Internal Medicine. Diseases of the Dog and Cat. Fifth Edition, Volume 2. Philadelphia: W.B. Saunders Company.

European Food Safety Authority (EFSA). (2008). Scientific Opinion. Mixture of chromium di- and tri-nicotinate as a source of chromium added for nutritional purposes in food supplements and in foods for particular nutritional uses. Scientific Opinion of the Panel on Food Additives and Nutrients Sources added to Food (ANS). The EFSA Journal. 887:1-24.

Expert Group on Vitamins and Minerals. (EVM). (2000). Review of Copper – Additional Cosideration of Animal Data. EVM/99/19/P. Accessed: January 9, 2012. Available at: http://www.food.gov.uk/multimedia/pdfs/evm9919p.pdf

Expert Group on Vitamins and Minerals. (EVM). (2002). Revised Review of Biotin. Accessed: January 7, 2012. Available at: http://www.food.gov.uk/multimedia/pdfs/ biotin.pdf

Flodin NW. (1990). Micronutrient supplements: toxicity and drug interactions. Progress in Food and Nutrition Science. 14(4):277-331.

Food and Nutrition Board (FNB), Institute of Medicine (IOB). (2001a). Dietary reference intakes for vitamin A, vitamin K, boron, chromium, copper, iodine, iron, manganese, molybdenum, nickel, silicon, vanadium, and zinc. Washington, D.C.: National Academy Press.

Food and Nutrition Board (FNB), Institute of Medicine (IOM). (2001b). Chromium. Dietary reference intakes for vitamin A, vitamin K, boron, chromium, copper, iodine, iron, manganese, molybdenum, nickel, silicon, vanadium, and zinc. Washington, D.C.: National Academy Press.

Friedrich W. (1988). Vitamins. Berlin, (FRG): Walter de Gruyter & Co.

Gerasimenko JV, Flowerdew SE, Voronina SG, et al. (2006). Bile acids induce Ca2+ release from both the endoplasmic reticulum and acidic intracellular calcium stores through activation of inositol trisphosphate receptors and ryanodine receptors. The Journal of Biological Chemistry. 281:40154-63.

Gladyshev VN. (2006). Selenoproteins and selenoproteomes. Accessed: January 2012. Available at: http://digitalcommons.unl.edu/cgi/viewcontent.cgi?article=1035&context=biochemgladyshev

Goodman & Gilman. (1996). The Pharmacological Basis of Therapeutics. Ninth Edition. Hardman JG, Limbird LE. (Eds). New York: McGraw Hill. pp. 1565.

Green AS, Tang G, Lango J, *et al.* (2011). Domestic cats convert [(2) H(8) ]-β-carotene to [(2) H(4) ]-retinol following a single oral dose. Journal of Animal Physiology and Animal Nutrition (Berl). doi: 10.1111/j.1439-0396.2011.01196.x.

Griffin C, Kwochka K, Macdonald J. (1993). Current Veterinary Dermatology: The Science and Art of Therapy. Mosby Publications. Linn, MO.

Groff JL, Gropper SS, Hunt SM. (1995). Advanced Nutrition and Human Metabolism. Second Edition. St. Paul (MN): West Publishing Company.

Haneke KE. (2002). Toxicological summary for sodium metasilicate and its pentahydrate and nonahydrate. Review of Toxicological Literature.

Integrated Laboratory Systems, Inc. North Carolina (USA). Accessed: January 9, 2012. Available at: http://ntp.niehs.nih.gov/ntp/htdocs/chem\_background/exsumpdf/ sodiummetasilicate.pdf

Hardy A, Krimer PM, Latimer KS. (2003). Canine Zinc Toxicosis. Veterinary Clinical Pathology Clerkship Program. Accessed: January 9, 2012. Available at: http://www.vet.uga.edu/vpp/clerk/hardy/

Hatch RC, Laflamme DP. (1989). Acute intraperitoneal cholecalciferol (vitamin D3) toxicosis in mice: its nature and treatment with diverse substances. Veterinary and Human Toxicology. 31:105-12.

Hatfield DL, Berry MJ, Gladyshev VN. (Eds). (2006). Selenium: Its molecular biology and role in human health. 2nd ed. New York: Springer.

Hendler SS, Rorvik DR, eds. (2001). PDR for Nutritional Supplements. Montvale: Medical Economics Company, Inc.

Jellin JM, Batz F, Hitchens K. (1999). Pharmacist's Letter/Prescriber's Letter Natural Medicines Comprehensive Database. 2nd ed. Stockton, CA: Therapeutic Research Faculty.

Johnson MA, Fischer JG, Kays SE. (1992). Is copper an antioxidant nutrient? Critical Reviews in Food Science and Nutrition. 32:1-31.

Kahn CM, Line S. (Eds). (2010). The Merck Veterinary Manual. 10th Edition. Whitehouse Station (NJ): Merck & Co., Inc.

Kasper H. (1999). Vitamin absorption in the elderly. International Journal of Vitamin and Nutrition Research. 69(3):169-172.

Khan KN, Andress JM, Smith PF. (1997). Toxicity of subacute intravenous manganese chloride administration in beagle dogs. Toxicologic Pathology. 25:344-50.

Kleiman DG, Thompson KV, Baer CK. (Eds). (2010). Wild Mammals in Captivity. Principles & Techniques for Zoo Management. 2nd Edition. Chicago (USA): The University of Chicago Press.

Klimis-Tavantzis DL. (Ed). (1994). Manganese in health and disease. Boca Raton: CRC Press, Inc.

Kukuljan M, Vergara L, Stojilkovic SS. (1997). Modulation of the kinetics of inositol 1,4,5-trisphosphate-induced [Ca2+]i oscillations by calcium entry in pituitary gonadotrophs. Biophysical Journal. 72(2 Pt 1):698-707.

Lamireau T, Bouchard G, Yousef IM, *et al.* (2007). Dietary lecithin protects against cholestatic liver disease in cholic acid-fed Abcb4- deficient mice. Pediatric Research. 61:185-90.

Larner J. (2002). D-chiro-inositol – its functional role in insulin action and its deficit in insulin resistance. International Journal of Experimental Diabetes Research. 3:47-60.

Leon AS, Spiegel HE, Thomas G, Abrams WB. (1971). Pyridoxine antagonism of levodopa in parkinsonism. JAMA. 218:1924-1927.

Lewis DP, Van Dyke DC, Willhite LA, Stumbo PJ, Berg MJ. (1995). Phenytoin-folic acid interaction. The Annals of Pharmacotherapy. 29(7-8):726-35.

Lewis LD. (2005). Feeding and Care of the Horse. Second Edition. Oxford, (UK): Blackwell Publishing.

Loflin J, Lopez N, Whanger PD, Kioussi C. (2006). Selenoprotein W during development and oxidative stress. Journal of Inorganic Biochemistry. 100:1679-84.

Lomaestro BM, Bailie GR. (1995). Absorption interactions with fluoroquinolones. Drug Safety. 12:314-33.

Massimino S, Kearns RJ, Loos KM, *et al.* (2003). Effects of age and dietary beta-carotene on immunological variables in dogs. Journal of Veterinary Internal Medicine. 17:835-42.

MDL Information Systems, Inc. (2002). Manganese Sulphate, OHS 13650. Nashville (TN). Accessed: January 9, 2012. Available at: http://www.michigan.gov/documents/ deq/deq-ess-lab-ManganeseSulfate\_316966\_7.pdf Meschino J. (2012). Meschino Health. Comprehensive Guide to Beta-Carotene. Accessed: February 10th, 2012. Available at: http://www.meschinohealth.com/books/ beta\_carotene.

Miranda DT, Batista VG, Grando FC, *et al.* (2008). Soy lecithin supplementation alters macrophage phagocytosis and lymphocyte response to concanavalin A: a study in alloxan-induced diabetic rats. Cell Biochemistry and Functions. 26:859-65.

Mochizuki M, Akagi K, Inoue K, Shimamura K. (1998). A single dose toxicity study of magnesium sulfate in rats and dogs (in Japanese). The Journal of Toxicological Sciences. 23 Suppl 1:31-5.

Moser LR, Smythe MA, Tisdale JE. (2000). The use of calcium salts in the prevention and management of verapamil-induced hypotension. The Annals of Pharmacotherapy. 34(5):622-629.

Mustacich D, Powis G. (2000). Thioredoxin reductase. The Biochemical Journal. 346 Pt 1:1-8.

Muszy ńska A, Pałka J, Gorodkiewicz E. (2000). The mechanism of daunorubicin-induced inhibition of prolidase activity in human skin fibroblasts and its implication to impaired collagen biosynthesis. Experimental and Toxicologic Pathology. 52:149-55.

National Research Council (NRC), Board on Agriculture (BOA). (1987). Vitamin Tolerance of Animals. Washington, D.C.: National Academy Press.

National Research Council (U.S.). (1986). Nutrient Requirements of Cats. National Academy Press, Wasington (DC).

National Research Council (U.S.). (2006). Nutrient Requirements of Dogs and Cats. Ad Hoc Committee on Dog and Cat Nutrition. National Academics Press.

National Research Council. (NRC). (2005). Mineral Tolerance of Animals. Second Revised Edition. The National Academies Press. Washington (DC).

Nielsen FH. Ultratrace minerals. In: Shils M, Olson JA, Shike M, Ross AC, Eds. (1999). Modern Nutrition in Health and Disease. 9th ed. Baltimore: Williams & Wilkins. 283-303.

O'Dell BL, Sunde RA. (Eds). (1997). Handbook of nutritionally essential minerals. New York: Marcel Dekker, Inc.

Parchure M, Ambaye RY, Lalitha VS, Gokhale SV. (1985). Acute toxicity of folic acid in mice. Experientia. 41:72-3.

Penttilä O, Hurme H, Neuvonen PJ. (1975). Effect of zinc sulphate on the absorption of tetracycline and doxycycline in man. European Journal of Clinical Pharmacology. 9:131-4.

Pfizer Inc. (2009). Material Safety Data Sheet. New York, NY. Accessed: January 10, 2012. Available at: http://www.pfizer.com/files/products/material\_safety\_data/ PZ00929.pdf

Pfizer Laboratories. (1990). Product Information: Vibramycin, doxycycline., New York, NY.

Phillips WE, Mills JH, Charbonneau SM, *et al.* (1978). Subacute toxicity of pyridoxine hydrochloride in the beagle dog. Toxicology and Applied Pharmacology. 44: 323-33.

Raila J, Rohn S, Schweigert FJ, Abraham G. (2011). Increased antioxidant capacity in the plasma of dogs after a single oral dosage of tocotrienols. British Journal of Nutrition. 106 Suppl 1: S116-9.

Rapiejko PJ, Northup JK, Evans T, et al. (1986). G-proteins of fat-cells. Role in hormonal regulation of intracellular inositol 1,4,5-trisphosphate. The Biochemical Journal. 240: 35-40.

Rayman MP. (2000). The importance of selenium to human health. Lancet. 356:233-241.

Robien K, Oppeneer SJ, Kelly JA, Hamilton-Reeves JM. (2013). Drug-Vitamin D Interactions: A Systematic Review of the Literature. Nutrition in Clinical Practice.

Schachter S, Nelson RW, Kirk CA. (2001). Oral chromium picolinate and control of glycemia in insulin-treated diabetic dogs. Journal of Veterinary Internal Medicine. 15:379-84.

Schenck P. (2010). Home-Prepared Dog & Cat Diets. Second Edition. Ames, Iowa, USA: Wiley-Blackwell.

Schulpis KH, Karikas GA, Tjamouranis J, Regoutas S, Tsakiris S. (2001). Low serum biotinidase activity in children with valproic acid monotherapy. Epilepsia. 42(10):1359-62.

Schweigert FJ, Raila J, Wichert B, Kienzle E. (2002). Cats absorb beta-carotene, but it is not converted to vitamin A. The Journal of Nutrition. 132:1610S-25.

Sciencelab.com, Inc. (2010a). Material Saftey Data Sheet. Calcium Carbonate. Houston, Texas. Accessed: January 8, 2012. Available at: http://www.sciencelab.com/msds.php?msdsId=9923249

Sciencelab.com, Inc. (2010b). Material Saftey Data Sheet. Inositol. Houston, Texas. Accessed: January 8, 2012. Available at: http://www.sciencelab.com/msds.php?msdsId=9923249

Shen X, Xiao H, Ranallo R, et al. (2003). Modulation of ATP-dependent chromatin-remodeling complexes by inositol polyphosphates. Science. 299:112-4.

Shils ME, Shike M, Ross AC, et al. (Eds). (2006). Modern Nutrition in Health and Disease. 10th ed. Philadelphia: Lippincott Williams & Wilkins.

Steger DJ, Haswell ES, Miller AL, et al. (2003). Regulation of chromatin remodelling by inositol polyphosphates. Science. 299: 114-6.

Sword JT, Pope AL, Hoekstra WG. (1991). Endotoxin and lipid peroxidation in vitro in selenium- and vitamin E-deficient and -adequate rat tissues. Journal of Nutrition. 121:258-264.

Tahiliani AG, Beinlich CJ. (1991). Pantothenic acid in health and disease. Vitamins and Hormones. 46:165-228.

Termanini B, Gibril F, Sutliff VE, Yu F, Venzon DJ, Jensen RT. (1998). Effect of long-term gastric acid suppressive therapy on serum vitamin B12 levels in patients with Zollinger-Ellison syndrome. The American Journal of Medicine. 104(5):422-430

Traber MG. (2008). Vitamin E and K interactions-a 50-year-old problem. Nutrition Reviews. 66(11):624-9. doi: 10.1111/j.1753-887.2008.00123.x.

Uauy R, Olivares M, Gonzalez M. (1998). Essentiality of copper in humans. The American Journal of Clinical Nutrition. 67(5 Suppl): 952S-959S.

University of Maryland Medical Center (UMMC). (2012a). Possible interactions with: Beta-Carotene. Accessed: February 10, 2012. Available at: http://www.umm.edu/ altmed/articles/beta-carotene-000941.htm

University of Maryland Medical Center (UMMC). (2012b). Possible interactions with: Vitamin B1 (Thiamine). Accessed: February 10, 2012. Available at: http://www.umm.edu/altmed/articles/vitamin-b1-000988.htm

University of Maryland Medical Center (UMMC). (2012c). Possible interactions with: Vitamin B2 (Riboflavin). Accessed: February 10, 2012. Available at: http://www.umm.edu/altmed/articles/vitamin-b2-000989.htm

University of Maryland Medical Center (UMMC). (2012d). Possible interactions with: Vitamin B9 (Folic Acid). Accessed: February 10, 2012. Available at: http://www.umm.edu/altmed/articles/vitamin-b9-000993.htm

University of Maryland Medical Center (UMMC). (2012e). Vitamin B12 (cobalamin). Accessed: February 10, 2012. Available at: http://www.umm.edu/altmed/articles/ vitamin-b12-000332.htm

University of Maryland Medical Center (UMMC). (2012f). Possible interactions with: Calcium. Accessed: February 10, 2012. Available at: http://www.umm.edu/altmed/articles/calcium-000945.htm

University of Maryland Medical Center (UMMC). (2012g). Possible interactions with: Magnesium. Accessed: February 10, 2012. Available at: http://www.umm.edu/ altmed/articles/magnesium-000968.htm

Unna K, Greslin JG. (1941). Studies on the Toxicity and Pharmacology of Pantothenic Acid. The Journal of Pharmacology and Experimental Therapeutics. 73:85-90.

USP Reference Standards. (2007). Material Safety Data Sheet. "Cyanocobalamin". The United States Pharmacopeial Convention, Inc. (2007). Catalog Number 1152009. Rockville, MD (USA). Accessed: January 8, 2012. Available at: umanitoba.ca/faculties/science/microbiology/MSDS/Cyanocobalamin\_VitB12.pdf

Vella A, Gerber TC, Hayes DL, Reeder GS. (1999). Digoxin, hypercalcaemia, and cardiac conduction. Postgraduate Medical Journal. 75(887):554-556.

Vieth R. (1999). Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. The American Journal of Clinical Nutrition. 69:842-56.

Watson TDG. (1998). Diet and Skin Disease in Dogs and Cats. The Journal of Nutrition. 128: 2783S-2789S.

Web MD. (2012). Beta-Carotene Interactions. Accessed: February 10, 2012. Available at: www.webmd.com/a-to-z-guides/common-topics/default.htm

Weir Jr. RJ, Fisher RS. (1999). Toxicologic studies on borax and boric acid. Encyclopedia of Food Microbiology: 1710-1717.

Wester PO. (1980). Urinary zinc excretion during treatment with different diuretics. Acta Medica Scandinavica. 208:209-12.

Yu S, Paetau-Robinson I. (2006). Dietary supplements of vitamins E and C and beta-carotene reduce oxidative stress in cats with renal insufficiency. Veterinary Research Communications. 30:403-13.

Zempleni J, Mock DM. (1999). Biotin biochemistry and human requirements. The Journal of Nutritional Biochemistry. 10:128-138.

Ziegler EE, Filer LJ. (Eds). (1996). Present Knowledge in Nutrition. 7th ed. Washington D.C.: ILSI Press.